<u>Statistical Process Control in Proteomics</u> SProCoP

This tutorial will guide you through the installation of SProCoP and using it to perform statistical analysis on a sample Skyline file.

Getting Started

To start this tutorial, download **SProCoPTutorial.zip** from this page:

https://skyline.gs.washington.edu/labkey/skyts/home/software/Skyline/tools/toolDetails.view?name=S ProCoP

Extract the files in it to a folder on your computer, like:

C:\Users\brendanx\Documents

This will create a new folder:

C:\Users\brendanx\Documents\SProCoPTutorial

The zip file contains the following files:

Skyline SProCoP.sky: A Skyline document containing sample data for SProCoP.

Skyline SProCoP.skyd: Contains extracted chromatogram data for the Skyline document.

Open the provided file Skyline_SProCoP.sky in Skyline. The peptides measured in the document along with the data acquisition parameters are described in **Figure 1** at the end of this tutorial. The data in the document are Quality controls that were monitored within the context of a larger study assessing the time course tryptic digestion of proteins from blood. All experimental data was collected on a Q-Exactive tandem mass spectrometer (ThermoFisher, Bremen, Germany) coupled to a nano Acquity UPLC (Waters). The bath gas in the C trap of the Q Exactive was low which led to decreasing ion abundance across the runs. The gas was then replenished and the targeted peak areas significantly increased.

Installing SProCoP as an External Tool

The Skyline external tools framework supports the direct integration of statistical analysis tools that process the data in a Skyline document.

To add SProCoP as an external tool, perform the following steps:

• On the **Tools** menu, click **External Tools** to bring up the **External Tools** form.

Menu contents:	
	Add
	Custom
	From File
	Move Up
	Move Down
Title:	
Command:	
Arguments:	
Initial directory:	
Input Report:	
	Output to Immediate Window

Note: Your External Tools menu may already contain some Tools

- Click the Add button, and select From File from the context menu.
- Navigate to the location of the SProCoP-1_0.zip file in the folder you created and select it.
- Click the **Open** button.

Skyline will now guide you through the process of installing SProCoP as an external tool. This may require you to install the R statistical programming environment, as well as a number of R packages which are used by SProCoP to perform its statistical analysis. For example, if you do not have the appropriate version of R installed, the following form will be displayed:

R Installer
This tool requires the use of R 3.0.2 and the following packages.
qcc
Install Cancel

To complete the installation, performing the following steps:

- On the **R Installer** form, click **Install**.
- If you do not have the specified version of R installed on your machine, wait for R to download. When the download is completed, you will be asked to install R onto your computer. Complete the R installation process by working through the R installer wizard.
- Once the R installation is completed, Skyline will download and install the necessary R packages for SProCoP. When prompted to run the "SkylineProcessRunner", click the **Yes** button to install packages.

Once the installation is complete, the SProCoP tool will appear in the **External Tools** form:

External Tools	×
Menu contents: SRM Collider QC_Rscript SProCoP	Add <u>R</u> emove
	Move Down
<u>T</u> itle:	SProCoP
<u>C</u> ommand:	\$(ProgramPath(R,3.0.2))
Arguments:	f "\$(ToolDir)\QCplotsRgui2.R"slaveno-s
Initial directory:	\$(DocumentDir)
Input Report: SProCoP Input	Output to Immediate Window OK Cancel Apply

• Click the **OK** button on the **External Tools** form.

Checking the Skyline Report

SProCoP will automatically generate a **SProCoP Input** report based on the .sky file. Although this step is not necessary we recommend that you check the Skyline Report to ensure that the report is rendered correctly. The demo Skyline_SProCoP.sky file report should look like the figure on the next page.

- On the File menu, click Export > Report.
- Select **SProCoP Input**, and click the **Preview** button on the right (see figure below)

Export Report	×
Report: Peptide Ratio Results SProCoPtutorial Peptide RT Results Transition Results SRM Collider Input SProCoP Input	<u>P</u> review <u>E</u> dit list <u>S</u> hare <u>Import</u>
Export	Cancel

Once **Preview** is selected, the **Preview Report** page will appear:

tu Pre	view Report							- • ×
	ReplicateName	PeptideModifiedSec	TotalArea	Best Retention Time	MaxFwhm	MinStartTime	MaxEndTime	AverageMassErrorPPM
•	Q_Exactive_08	VYVEELKPTPE	110572224512	29.55	0.1	29.41	29.76	0.8
	Q_Exactive_08	VYVEELKPTPE	160706428928	29.5	0.14	29.41	29.78	1.1
	Q_Exactive_08	VYVEELKPTPE	100025868288	46.42	0.19	46.29	46.93	1
	Q_Exactive_08	VYVEELKPTPE	173198966784	29.39	0.18	29.31	29.68	1.8
	Q_Exactive_08	VYVEELKPTPE	153556402176	29.36	0.16	29.26	29.66	1
	Q_Exactive_08	VYVEELKPTPE	176071475200	29.32	0.22	29.24	29.63	1.2
	Q_Exactive_08	VYVEELKPTPE	148374831104	29.39	0.15	29.26	29.65	1.3
	Q_Exactive_08	VYVEELKPTPE	137785720832	29.42	0.1	29.34	29.73	0.8
	Q_Exactive_08	VYVEELKPTPE	166037176320	29.49	0.17	29.31	29.68	1.6
	Q_Exactive_08	VYVEELKPTPE	145370038272	29.42	0.17	29.29	29.73	0.8
	Q_Exactive_08	VYVEELKPTPE	142750416896	29.36	0.17	29.28	29.68	0.9
	Q_Exactive_08	VYVEELKPTPE	155523088384	29.44	0.13	29.31	29.68	0.5
	Q_Exactive_08	VYVEELKPTPE	115039272960	29.41	0.16	29.32	29.71	0.9
	Q_Exactive_08	VYVEELKPTPE	112991633408	29.41	0.16	29.31	29.7	1.6
	Q_Exactive_08	VYVEELKPTPE	107816869888	29.47	0.18	29.34	29.7	1.2
	Q_Exactive_08	VYVEELKPTPE	128046080000	29.36	0.11	29.28	29.7	1.1
	Q_Exactive_08	VYVEELKPTPE	130433499136	29.37	0.1	29.26	29.65	1.2
	Q_Exactive_08	VYVEELKPTPE	116482187264	29.45	0.15	29.29	29.66	2.2
	Q_Exactive_08	VYVEELKPTPE	127296757760	29.37	0.13	29.28	29.6	1.9
	Q_Exactive_08	VYVEELKPTPE	106591420416	29.44	0.11	29.34	29.7	2.6
	Q_Exactive_08	VYVEELKPTPE	96447987712	29.39	0.11	29.29	29.63	2.5
	Q_Exactive_08	VYVEELKPTPE	91348934656	29.44	0.13	29.29	29.65	2.4
	Q_Exactive_08	VYVEELKPTPE	82229624832	29.34	0.17	29.26	29.65	3.4
	Q Exactive 08	VYVEEI KPTPE	72177934336	29.39	0 07	29 29	29.62	35 *
* ''' Record: I 1 of 329 > Filter: '''								

Using SProCoP

To run SProCoP, perform the following steps:

• On the **Tools** menu, click **SProCoP**.

Skyline will begin exporting the SProCoP input report. This may take a few minutes. You can track the progress of the report export with the green progress bar in the bottom of the Skyline Window. Once the report is exported, Skyline will display the **SProCoP** form:

SProCoP
<u>N</u> umber of runs to establish thresholds: β →
 Using high resolution instrument Absolute mass measurement accuracy: ppm
Save as PDF
OK Cancel

The "Using high resolution instrument" checkbox should be checked if your data was acquired on a high resolution instrument. The reports generated by SProCoP can be saved to a PDF file by checking the **Save as PDF** checkbox. High quality charts are saved to a single PDF file with one chart per page.

- For this tutorial you will leave the options in their default settings.
- Click on the **OK** button.

SProCoP will generate two windows (pictured below). The larger window represents a control chart matrix for all the targeted peptides across 4 different metrics: Peak areas, RT, FWHM and Peak asymmetry. The first three are all reported by Skyline directly from the raw data. Peak asymmetry is calculated by Pasym=(a+b)/2a and determined from the min start, RT at apex, and max end. Parameters a and b are the width at base from the start of the peak to the apex and from the apex to the end of the peak, respectively. For calculation of MMA, a MMA value is calculated and stored for every time-intensity pair in the chromatograms it extracts from high resolution data. The mass error at each chromatogram point is calculated by taking a weighted average of the m/z-intensity pairs found within the m/z extraction window, and subtracting the expected m/z from that average. When Skyline performs linear interpolation of extracted chromatogram points onto a scale with a consistent interval, it applies the same interpolation to the mass error values. Finally, to calculate the mass error for any integrated peak, Skyline calculates a weighted average of the mass errors for chromatogram points within the integration boundaries, weighted by their background-subtracted intensities. For more information on the control chart see **Figure 2** at the end of the tutorial.

The smaller window represents a Pareto Chart which is a combination of a bar and line graph. The bar graph represents the frequency of times a peptide, across all the runs, is outside the \pm 3sd threshold determined empirically for the 4 metrics. The line graph is a cumulative percentage chart and represents the proportion of total "nonconformers" (i.e., outside \pm 3 sd) that a particular metric constitutes.



Figure 1

Parameter	Setting
Source	Nano-ESI
Polarity	Positive
Capillary temperature (°C)	325°
S-Lens RF Voltage	50%
Source Voltage (kV)	2.2

Full-scan MS parameter	Setting
Mass range (m/z)	400-2000
Resolution (FWHM at <i>m</i> /z 200)	70,000
AGC Target	1 x 10 ⁶
Spectrum data type	Profile
Maximum injection time (msec)	10

Targeted MS/MS parameters	Setting
Isolation window (<i>m/z</i>)	2
Resolution (FWHM at <i>m/z</i> 200)	17,500
AGC Target	1 x 10 ⁶
Spectrum data type	Profile
Collision energy (NCE)	25
Default charge state	+2
Maximum injection time (msec)	55

Peptide from bovine QC mix	Mono-isotopic mass for 2+	Retention Time
	precursor ion	
VLVLDTDYK	533.29500	23.2
VLDALDSIK	487.28190	23.8
ATEEQLK	409.71630	14.5
DDGSWEVIEGYR	713.31770	26.0
YSTDVSVDEVK	621.29840	20.0
FFVAPFPEVFGK	692.86860	30.8
VYVEELKPTPEGDLEILLQK	1157.13300	28.0
LVNELTEFAK	582.31900	24.6

Figure 2

A detailed description of the control chart used in the studies. The blue line represents the empirically defined mean of the process and the control lines are 1s, 2s, and 3s from the mean. The control lines are determined empirically from the user specified number of QC standards. The CV% represents the coefficient of variation across all runs. It's the standard deviation of all QC runs divided by the mean of all QC runs. In this particular example 46 standards were used to calculate the CV shown.

